**GROWTH RESPONSE, CARCASS AND HAEMATOLOGICAL PARAMETERS OF BROILER BIRDS FED SELECTED MEDICINAL PLANTS LEAF MEAL.**

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**Abstract**

One hundred and twenty (120) healthy day-old Ross 308 broiler chicks, were used in a study to evaluate the effects of substitution of antibiotics with selected medicinal plant leaf meal on their production indices. The study which was carried out from September to November 2022 consists of eight treatment groups designated as T1 to T8. Eight experimental diets were formulated such that T1 served as the control, T2 (5% *Moringa oleifera* leaf meal), T3 (5% Neem leaf meal) T4 (5% *Spondias mombin* leaf meal), T5 (2.5% combination each of moringa and Neem leaf meal), T6 (2.5% combination each of *Moringa* and *Spondias mombin* leaf meal) T7 (2.5% combination each of Neem and *Spondias mombin* leaf meal), and T8 (1.66% each of *Moringa*, Neem and *Spondias mombin* leaf meal) respectively. Parameters evaluated for the broiler chicken include weight gain, feed intake, feed conversion ratio, mortality, haematology and carcass characteristics of the broilers. The study lasted for 63 days. Data collected were subjected to a one-way analysis of variance using SPSS,22. Mean separation was done using the least significant difference. The weight gain, feed intake, feed conversion ratio as well percentage mortality for the broiler chicken all differed (P<0.05). The broiler chicken dressed weight, dressing percentage, breast cut, thigh, drumstick, shank and wing were significant (P<0.05) higher for T4. The organs of the broiler chicken such as gizzard, crops, proventriculus, small intestine, large intestine and caecum were not significantly (P>0.05) affected by the feeding trial. The WBC, Hb and the platelet of the broiler chicken were significantly (P<0.05) affected by the study materials. The work therefore recommends that medicinal plant leaf meal can be used in place of antibiotics growth promoters as they positively affected the broiler growth performance.

**Key words: Medicinal plant, broiler starter, broiler finisher, growth rate, dressing percentage, Blood parameters**

**Introduction**

Alternative ingredients are being researched to replace all or some conventional foods with alternative diets made from roots and tubers like cassava and new legumes such the jack bean and the sword bean (Iji 1999). This is true everywhere in the world, but is especially true in regions where there is a feed shortage. Due to nutrient inadequacies, such as those in amino acids, mineral imbalances in energy protein ratios, and anti-nutritive substances including non-starch polysaccharides (NSPs), polyphenols, and phytic acid, poultry productivity is frequently low (Dilger et al., 2008).

In order to enhance the quality of alternative diets, chicken feed manufacturers employ a number of techniques, such as nutrient addition and feed processing. According to a review of the usage of feed additives by Iji et al. (2011), additives are used to raise the quality of poultry meals. These additives are chemicals that are added in minuscule amounts (Alaku, 2010). According to Ugwu (2006), they also include certain amino acids (lysine and methionine), hormones, arsenic, tranquillizers, and antioxidants.

Minerals and vitamins, in addition to additives and supplements, are now used in feeds (Sonaiya, 1993). However, the Food and Drug Administration (FDA) of the United States of America (USA) and the majority of European countries, including Sweden, Belgium, Germany, and others, have discouraged the use of chemicals, particularly antibiotics, in livestock feeds (Customer Updates, 2013). This is due to the persistence of the effects in cattle products and the emergence of drug-resistant microorganism strains (Oyekunle and Owonikoko, 2002). Currently, the use of feed additives is prohibited in the majority of developed nations (Ugwu, 2006).

As a result, additives with organic and eco-friendly qualities are sought for (Adedeji et al., 2008). Some of these compounds, primarily those with a botanical origin, have already been used. To cure ectoparasites in chicken, for instance, Carica papaya leaf can be burned to ashes and applied topically (Nwude and Ibrahim, 1980). According to Maigandi and Usman (1996), turkeys with coughing and diarrhoea can be treated with Zingiber officinale (ginger). According to Gefu et al. (2000), citrus aurantifolia is utilised in chicken species to prevent worm infestation. Elaeis guinensis (oil) is a plant that is used to cure ectoparasites in all species of poultry, according to Adedeji et al. (2013).

Herbs (medical plants) are often derived from plants, and plants create certain metabolites known as phytochemicals as part of their biological processes. According to DalleZotte et al. (2016), phytochemicals can be categorized depending on their therapeutic qualities, such as antibacterial, antifungal, anti-inflammatory, antiulcer, antioxidant, antiviral, anticancer, and immunological stimulants. Given that there are around 500,000 medicinal plants in the globe and that many of their therapeutic characteristics have not been researched, medicinal plants represent a potential source of medicine with a bright future. About 21,000 plant species, according to FAO (2008), have the potential to be utilized as medicines and are regarded as safe since they have either no adverse effects or only minor side effects.

The purpose of this study, therefore, is to investigate the effect of substituting antibiotics with three selected medicinal plant leaf meals (*Moringa oleifera, Azadirachta indica, Spondias mombin* and their combinations) on the growth performance, carcass characteristics and blood parameters of the broiler birds.

**Materials and method**

**Description of the study area.**

A field experiments were conducted at the Poultry Unit, Department of Animal/Fisheries Science and Management, Teaching and Research Farm of the Faculty of Agriculture and Natural Resources Management, Enugu State University of Science and Technology, (ESUT) Agbani. The location is 670 4 North latitude, 803 East longitude, and 450 metres above sea level (Anikwe *et al*., 2017). The yearly rainfall in the area ranges from 1700 to 2010mm. The climate pattern is bimodal, with a rainy season from April to October and a dry season from November to March. The textural class of the soil is sandy loam with an isohyperature regime (Anikwe *et al*, 2017), and it has been classed as Typical paleudults of the Order ultisol (Anikwe *et al*.,2016).

**Experimental materials and Preparation**

The *Moringa oleifera* (drum stick) and *Azadirachta indica* leaves(neem) were collected from ESUT commercial farm premises while *Spondias mombin* leaf was collected from Umueze in Nkanu West Local Government Area, Enugu State. The leaves were dried on a well-cleaned cemented floor. They were evenly spread and regularly turned to encourage fast and even drying. To make each leaf meal, the leaves were milled separately using a hammer mill when they were crispy while still retaining the greenish colouration. Various leaf samples were analysed to determine their proximate composition, mineral profile, and phytochemical composition. The analysis was conducted at the Animal Science Laboratory, University of Nigeria, Nsukka.

**Design of the experiment**

In a randomised block design, day-old chicks were randomly assigned to one of eight feed treatments based on their weight. T1, (the control); T2, (5% MOLM), T3, (5% AILM), T4, (5% SMLM), T5, (2.5% each of MOLM + AILM), T6, (2.5% each of MOLM + SMLM), T7, (2.5% each of AILM +SMLM) and T8, (1.66% of MOLIM + AILM + SMLM). Each treatment contained eighteen (15) birds. Each treatment was repeated three times, with each replicate including five birds. The birds were rigorously cared for, with *ad libitum* feed and water. The study lasted for 63 days.

MOLM *Moringa oleifera* leaf meals

 AILM *Azadirachta indica* leaf meals

SMLM *Spondias mombin* leaf meals.

**Experimental house.**

This investigation was conducted in one of the chicken houses of Enugu State University of Science and Technology's Teaching and Research Farm, Poultry Unit. The poultry house has dwarf walls and is completely netted to allow for proper ventilation. A deep litter system was used to handle the birds. On both sides of the poultry house, 24 apartments measuring 1.8m x 1.5m and 0.7-0.9m2 floor area per bird were created. The pens were divided with half-inch wire mesh and wood. One treatment duplicate was kept in each pen. The litter materials were cleaned out every two weeks to maintain the sanitary state of the pens.

**Experimental birds and management.**

One hundred and twenty (120) healthy day-old Ross 708 broilers were purchased from Avian Chicks, Uwani, Enugu. The deep litter system was adopted. The house was washed and disinfected before the chicks arrived. The brooding of the chicks lasted three (3) weeks. Two 100-watt electrical lights were kept at roughly 15cm above the ground during brooding to provide heat for each pen, then gradually raised to 1.75m height towards the conclusion of the brooding. Kerosene burners and charcoal heaters were utilised as backup heat sources during brooding if the electricity went out. The temperature during brooding was maintained at 390C from the first day to the second day and this will be gradually reduced by 20C every week to a final temperature of 33.5oC till the end of brooding on the 21st day. Routine management practices, including hygiene, vaccination, and medication, were observed.

**Experimental Design.**

In a randomised block design, day-old chicks were randomly assigned to one of eight feed treatments based on their weight. T1, (the control); T2, (5% MOLM), T3, (5% AILM), T4, (5% SMLM), T5, (2.5% each of MOLM + AILM), T6, (2.5% each of MOLM + SMLM), T7, (2.5% each of AILM +SMLM) and T8, (1.66% of MOLIM + AILM + SMLM). Each treatment contained eighteen (15) birds. Each treatment was repeated three times, with each replicate including five birds. The birds were rigorously cared for, with *ad libitum* feed and water. The study lasted for 63 days.

**Experimental procedure**

Eight experimental diets were formulated for the broiler birds. The eight experimental diets are T1, (control), T2, (5% MOLM) T3, (5% AILM) T4, 5% SMLM) T5, (2.5% each of MOLM +AILM) T6, (2.5% each of MOLM +SMLM) T7 (2.5% each of AILM + SMLM) and T8 (1.66% each of MOLM+ AILM + SMLM.

**Experimental diet**

Eight experimental diets were formulated; one for each treatment group. The starter diet was fed from 0 – 4 weeks, while the finisher’s diet was fed from 5 –9 weeks. The composition of the experimental diets is shown in Tables 1 and 2;

**Table 1; Composition of the broiler Starter experimental diet (100kg)**

|  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| Ingredients | T1 | T2 | TREATMENTS T3 | T4 | T5 | T6 | T7 | T8 |  |  |
| Maize | 53.00 | 53.00 |  53.00 | 53.00 | 53.00 | 53.00 | 53.00 | 53.00 |  |  |
| Groundnut cake | 14.00 | 14.00 |  14.00 | 14.00 | 14.00 | 14.00 | 14.00 | 14.00 |  |  |
| Soya bean | 20.00 | 20.00 |  20.00 | 20.00 | 20.00 | 20.00 | 20.00 | 20.00 |  |  |
| Fish meals | 4.00 | 4.00 |  4.00 | 4.00 | 4.00 | 4.00 | 4.00 | 4.00 |  |  |
| Wheat offal | 4.97 | 5.00 |  5.00 | 5.00 | 5.00 | 5.00 | 5.00 | 5.00 |  |  |
| Oxytetracycline\*  | 0.03 | - |  - | - | - | - | - | - |  |  |
| MOLM | - | 5.00 |  - | - | 2.5 | 2.5 | - | 1.66 |  |  |
| AILM | - | - |  5.00 | - | 2.5 | - | 2.5 | 1.66 |  |  |
| SMLM | - | - |  - | 5.00 | - | 2.5 | 2.5 | 1.66 |  |  |
| Limestone | 2.00 | 2.00 |  2.00 | 2.00 | 2.00 | 2.00 | 2.00 | 2.00 |  |  |
| Bone meals | 1.00 | 1.00 |  1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 |  |  |
| Salt | 0.25 | 0.25 |  0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 |  |  |
| Lysine | 0.25 | 0.25 |  0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 |  |  |
| Methionine | 0.25 | 0.25 |  0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 |  |  |
| PremixTotalCal. Analysis | 0.25100kg | 0.25100kg |  0.25 100kg | 0.25100kg | 0.25100kg | 0.25100kg | 0.25100kg | 0.25100kg |  |  |
| Crude Protein (%) | 24.75 | 23.89 |  23.55 | 22.62 | 23.86 | 22.35 | 23.88 | 22.65 |  |  |
| Metab energy (kcal/kg) | 2800 | 2868 |  2743 | 2754 | 2813 | 2853 | 2766 | 2688 |  |  |
| Crude fibre (%) | 3.44 | 3.54 |  3.64 | 3.88 | 3.23 | 3.65 | 3.32 | 3.93 |  |  |
| Calcium (%) | 1.48 | 1.52 | 1.35 | 1.65 | 1.73 | 1.87 | 1.39 | 1.68 |  |  |
| Av Phos (%) | 0.68 | 0.88 | 0.69 | 0.87 | 0.68 | 0.73 | 0.58 | 0.93 |  |  |
|  |  |  |  |  |  |  |  |  |  |  |

\*30g/100kg of feed non-therapeutic use

**Table 2: Composition of broiler Finisher experimental diet (100kg)**

|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
| Ingredients | T1 | T2 | T3 | TreatmentsT4 | T5 | T6 | T7 | T8 |
| Maize | 54 | 54 | 54 | 54 | 54 | 54 | 54 | 54 |
| Groundnut cake | 18.20 | 18.20 | 18.20 | 18.20 | 18.20 | 18.20 | 18.20 | 18.20 |
| Soya bean | 17.70 | 17.70 | 17.70 | 17.70 | 17.70 | 17.70 | 17.70 | 17.70 |
| Fish meals | 3.00 | 3.00 | 3.00 | 3.00 | 3.00 | 3.00 | 3.00 | 3.00 |
| Wheat offal | 9.97 | 5.00 | 5.00 | 5.00 | 5.00 | 5.00 | 5.00 | 5.00 |
| Oxytetracycline\* | 0.03 | - | - | - | - | - | - | - |
| MOLM | - | 5.00 | - | - | 2.50 | 2.50 |  | 1.66 |
| AILM | - | - | 5.00 | - | 2.50 | - | 2.50 | 1.66 |
| SMLM | - | - | - | 5.00 | - | 2.50 | 2.50 | 1.66 |
| Limestone | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 |
| Bone meals | 2.00 | 2.00 | 2.00 | 2.00 | 2.00 | 2.00 | 2.00 | 2.00 |
| Salt | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 |
| Lysine | 0.10 | 0.10 | 0.10 | 0.10 | 0.10 | 0.10 | 0.10 | 0.10 |
| Methionine | 0.20 | 0.20 | 0.20 | 0.20 | 0.20 | 0.20 | 0.20 | 0.20 |
| Premix | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 |
| TotalCal. Analysis | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 |
| Crude protein (%) | 18.08 | 18.81 | 18.18 | 18.21 | 18.36 | 18.16 | 18.22 | 18.19 |
| Metab energy(kcal/kg) | 3023 | 3037 | 3107 | 3076 | 3093 | 3084 | 3062 | 3058 |
| Crude fibre (%) | 4.25 | 4.60 | 4.36 | 4.30 | 4.48 | 4.63 | 4.59 | 4.63 |
| Calcium (%) | 1.46 | 1.50 | 1.48 | 1.48 | 1.46 | 1.48 | 1.47 | 1.48 |
| Phos(%) |  0.66 | 0.68 | 0.69 | 0.71 | 0.74 | 0.72 | 0.69 | 0.68 |
|  |  |  |  |  |  |  |  |  |

\*30g/100kg of feed for non-therapeutic use**.**

**Performance evaluation of broiler chicken**

Data on body weight increase, feed intake, and mortality data were collected, and the feed conversion ratio (FCR) was calculated.

**Average weight gain (g)/bird**

The birds were weighed at the start of the experiment and then every week after that.

 Average weight gain = weight of birds (g) - Initial weight (g)

 Total of birds

**feed intake (g)/bird**

The birds were given a known amount of feed, and the leftover feed was weighted to establish average daily and weekly feed intake. The feed intake was determined using the following formula:

 Average feed intake per bird = feed supplied (g)- leftover of feed (g)

 Number of birds

**Feed conversion ratio (FCR)**

The FCR of each group of birds was computed by dividing feed intake by body weight gain and was thus calculated as:

Feed conversion ratio (FCR) = Total feed intake (g)/bird

 Total body weight gain (g)/bird

**Mortality percentage=** Number of dead birds x 100

 Number of birds

**Determination of** **haematological parameters**

At the end of the experiment, six birds from each group were selected for collection of blood samples from the wing vein. For estimation of haematological parameters blood was collected aseptically with anticoagulant and estimated using an automatic haematolyzer. For estimation of total serum biochemical profile blood was collected aseptically from the birds. The blood samples were brought to the laboratory without disturbing the clots and centrifuged at 3000 rpm for 15 minutes to collect serum and stored at -20°C till further analysis.

**Determination of microbial counts in faecal samples culture media**

The good selective media used include the MacConkey agar media (Fluka, UK), *Salmonella Shigella* agar (BIOTECH Laboratories LTD, UK), Citrate agar (BIOTECH Laboratories LTD, UK), and Peptone water (BIOTECH Laboratories LTD, UK) (Oxoid, UK). Following the manufacturer's instructions, all culture medium was prepared aseptically.

**Analyses of animal droppings:** With minor modifications, this was carried out according to Okafor and Umeh's (2013) techniques. Faecal samples (1g from each animal dropping) were homogenised in sterile distilled water, to summarise. The resultant suspensions were sorted and serially diluted by transferring 1 mL of each suspension into a corresponding sterile test tube containing 9 mL (10-1) of sterile distilled water. The diluted homogenates were then aseptically plated on MacConkey agar plate 10-1 and Salmonella shigella agar plate 10-1, respectively, using a sterile pipette. A sterile glass spreader was used to apply this. The agar plates might be dried, then incubated for 48 hours at 37 degrees Celsius.

**Statistical analysis.**

Using Speciial Package for Social Science (SPSS) statistical software, version 22.00 for Windows, all data were subjected to one-way analysis of variance (ANOVA) (SPSS, 2012). Where statistical differences were found between means, they were separated using the least significant differences (LSD) procedure at a significance level of 5% as contained in the same statistical package.

**Results and discussion**

Effects of substitution of antibiotics with graded levels of selected medicinal plant leaf meals on the growth performance of broiler chicken are presented in Table 3;

**Table 3: Effects of substitution of antibiotics with selected medicinal plant leaf**

**meals on broiler starter performance**

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Parameters** | **1** | **2** | **3** | **TREATMENTS****4** | **5** | **6** | **7** | **8** | **SEM +** |
| Av initial wgt(g) | 41.2 | 40.7 | 42.06 | 39.76 | 40.73 | 41.73 | 42.13 | 40.02NS | 0.83 |
| Av final wgt(g)AV body wt gain(g) | 969b927.8 | 1003a962.7  | 965b922.4 | 1057a1017.24 | 995ab954.27 | 999ab956.87 | 1038a995.87 | 1025a984.98 | 2.451.83 |
| Avdaily feed intake(g) | 36.64b | 36.61b | 33.63c | 42.94a | 34.84c | 34.95c | 35.78b | 37.13b | 0.13 |
| Avdaily wgt gain(g) | 33.13 | 34.3b | 26.37 | 36.33 | 34.08 | 34.19 | 35.57 | 35.18NS | 0.63 |
| FCR | 1.10b | 1.06a | 1.27b | 1.18b | 1.02a | 1.02a | 1.01a | 1.05a | 0.06 |
| Mortality  | 5.55b | - | 11.11a | - | - | - | - | - | 0.13 |

Means within a row with different superscripts differ (P<0.05) significantly

**Weight gain**

The result of the feeding trial showed that at the end of the starter phase, the final body weight was highest for T4 (1057) g, followed by T7 (1038) g, T8 (1025) g, T2 (1003) g, T6(999) g, T5(995) g, T1 (969) g and T3 (965) g. The result showed that the body weight of birds in T4, T7, T8 and T2, which contain the selected medicinal plant leaf meals, was significantly (P<0.05) higher than those in other treatment groups. The superior significant (P<0.05) performance of T4 over the other treatment groups may be due to the rich phytochemical content of T4 which improved gut function and dietary palatability (Frankic *et al*., 2009). The final body weight of the broilers in T4 (SMLM) has also been attributed to minerals, vitamins and phytochemical present in *Spondias mombin* leaf meals which has a higher biological function which acted as a growth promoter, absorption enhancers, antimicrobial agents, and metabolic modifiers (Gill, 2001; Abaza, 2001; Hassan, 2010).

**Feed intake**

The average daily feed intake for the treatment groups during the starter phase indicated that T4 birds consumed more feed (42.94) g followed by T8 (37.13) g, T1 (36.64) g, T2 (36.61) g, T7 (35.78) g, T6 (34.95) g, T5 (34.84) g and T3 (33.63) gram respectively. Thus, the average daily feed intake during the starter phase was significantly (P<0.05) high for T4 and low for T3. The significant (P>0.05) reduction in feed intake shown by T3 at this phase may be because of the bitter taste of neem leaf meals, high fibre content and high oxalate content of the leaf meals of neem which may have resulted in low feed intake and, thus the poor performance of the birds fed neem leaf meal (T3)

**Feed conversion ratio**

The feed conversion ratio during the first four weeks (starter stage) was best for T7 (1.01) followed by T5 (1.02) and T6 (1.02) T8 (1.05), T2 (1.06), T1 (1.10), T4 (1.18) and T3 (1.27) in that order. Though numerical differences were observed during this phase, there was no significant (P>0.05) difference among the treatment groups.

The best feed conversion ratio shown by T4 birds that consumed *Spondias mombin* was due to the phytochemical content of *Spondias mombin* which lowered the pH of the digestive organ resulting in better utilization of nutrients (Seema and Johri, 1992; Bengmark, 1998; Dhama, 2011). The antimicrobial content of *Spondias mombin* changed the intestinal microflora, which helped to improve broiler performance, health status and reduced the microbial use of nutrients (Snyder and Wostmann, 1987). Lowering the intestinal pH by the phytochemical present in T4 also optimized the activity of protease and beneficial bacterial (Overland *et al*., 2000; Nava *et al*., 2009) and thus enhanced better feed conversion by the birds.

**Mortality rate**

The feeding trial showed that during the starter phase, the highest mortality was recorded in T3 (11.11 percent) followed by T1 (5.55 percent) mortality. The other treatment groups recorded no mortality. The bio-pesticidal agent in neem besides the disulphide is mainly azadirachtin and salannin, which are toxic. Thus, they may have been responsible for the death recorded in the first week of the life of the broiler chicken fed neem leaf meals. The death of 5.55 percent in the control group (T1) may be most probably because of resistance to antibiotics.

**Table 4:****Effects** **of substitution of antibiotics with selected medicinal plant leaf meals**

**on finisher broiler** **performance**

|  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  **TREATMENTS**

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Parameters** |  **1** |  **2** |  **3** |  **4** |  **5** |  **6** |  **7** |  **8** | **SEM+** |
| Av initial wgt (g) | 969 | 1003 | 965 | 1057 | 995 | 999 | 1038 | 1025 | 0.83 |
| Av final wgt (g) | 2988b | 3055ab | 2729b | 3329a | 3011ab | 3023ab | 3128a | 3112a | 8.23 |
| Av daily feed intake(g) | 188.33a | 187.81ab | 177.9c | 182.57ab | 183.71ab | 184.90ab | 181.58ab | 191.51a | 8.64 |
| Av daily wgt gain(g) | 56.51 | 57.4b | 49.20 | 63.78 | 56.44 | 6.64 | 58.51 | 58.45 | 4.03 |
| FCR | 3.33ab | 3.27ab | 3.61b | 2.86a | 3.25ab | 3.26ab | 3.10ab | 3.27ab | 0.52 |
| Mortality (%) | - | - | - | - | - | - | - | - | - |

 |  |  |  |  |  |  |  |  |  |

Means within a row with different superscripts differ (P<0.05) significantly

**Weight gain**

The finisher phase showed T4 (SMLM) again had the highest weight of 3329g, closely followed by T7 (3128) g, T8 (3112) g, T2 (3055) g, T6 (3023) g, T5 (3011) g, T1 (2988) g and T3 (2729) g, respectively. The average final weight for T4, T7 and T8 was significantly (P<0.05) higher than for the other treatment groups. The superior significant (P<0.05) performance of T4 over the other treatment groups may be due to the rich phytochemical content of T4, which improved gut function and dietary palatability (Frankic *et al*., 2009). The final body weight by the broilers in T4 has also been attributed to minerals, vitamins, and phytochemicals present in *Spondias mombin* leaf meals which have a higher biological function and acted as a growth promoter, absorption enhancers, antimicrobial agents and metabolic modifiers (Gill, 2001; Abaza, 2001; Hassan *et al*., 2010).

**Feed intake**

The average daily feed intake for the broiler’s finisher phase was highest for T8 (191.51) g followed by T1 (188.33) g, T2 (187.81) g, T6 (184.90) g, T5 (183.71) g, T4 (182.57) g, T7 (181.58) g and T3 (177.90) g respectively. Significant (P<0.05) differences were also observed among the treatment groups. Hence, the feed intake of T3 was significantly (P>0.05) lower when compared to other treatment groups. The significant (P<0.05) increased feed intake shown by T4 during the brooding phase may be because of the high biological function, mineral and vitamin and its low tannins content of *Spondia mombin* which helps to enhance appetite.

The significant (P>0.05) reduction in feed intake shown by T3 at the finisher phase may be because of the bitter taste of neem leaf meals, high fibre content and high oxalate content of the leaf meals of neem which may have resulted in low feed intake and, thus the poor performance of the birds fed neem leaf meals (T3). Low feed intake by the treatment group containing neem leaf could also result from the anti-feeding properties because of its efficacy in suppressing feed intake, even at a concentration of less than one part per million (Isman *et al*.,1991).

**Feed conversion ratio**

During the finisher phase, however, the results showed that the feed conversion ratio was best for T4 (2.86) followed by T7 (3.10), T5 (3.25), T6 (3.26), T2 and T8 (3.27), T1(3.33) and T3 (3.61) respectively. Broiler birds in T4 during the same period significantly (P<0.05) had the best FCR compared with broilers in other treatment groups. It was also observed from the feeding trial that broiler birds in T3 significantly (P>0.05) converted the lowest feed to body tissues during the finisher phase.

The best feed conversion ratio shown by T4 birds, that consumed *Spondias mombin* was because of the phytochemical content of *Spondias mombin* which lowered the pH of the digestive organ resulting in better utilization of nutrients (Seema and Johri, 1992; Bengmark, 1998; Dhama, 2011). The antimicrobial content of *Spondias mombin* may also have changed the intestinal microflora, which helped to improve broiler performance, health status and reduced the microbial use of nutrients (Snyder and Wostmann, 1987). Lowering the intestinal pH by the phytochemicals present in T4 also optimised the activity of protease and beneficial bacterial (Pranen and Morz, 1999; Overland *et al*., 2000; Nava *et al*., 2009) and thus enhanced better feed conversion by the birds.

**Mortality rate**

However, during the broiler finishing phase, no mortality was recorded. This may be because the birds at this phase no longer react to the bio-pesticidal agent in neem, mainly disulphide, azadirachtin and salannin which are insecticidal. These may have been responsible for the death recorded in the first week of the life of the broiler chicken fed neem leaf meals.

The effect of substitution of antibiotics with selected medicinal plant leaf meals on carcass

characteristics of broiler chicken are presented in Table 5;

**Table 5: Effects of the selected medicinal plant leaf meals on carcass characteristics broiler**

|  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  **TREATMENT**

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Parameters** |  **1** |  **2** |  **3** |  **4** |  **5** |  **6** |  **7** |  **8** | **SEM +** |
| Av final wgt (g)  | 2988b | 3055ab | 2729b | 3329a | 3011ab | 3023ab | 3128a | 3112a | 8.23 |
| Dressing (%) | 65.57b | 67.64ab | 64.432b | 72.17a | 65.67b | 67.37ab | 68.23ab | 67.77ab | 1.38 |
| Breast cut(g) | 389.38b | 398.46b | 331.71ab | 511a | 409.76ab | 405.76ab | 362.2b | 374.32b | 14.74 |
| Thigh (g) | 289.64ab | 304.21ab | 250.94c | 357.38a | 270.51c | 350.32a | 289.03b | 292.74ab | 11.13 |
| Drumstick (g) | 273.78c | 293.36b | 245.22c | 460.09a | 270.83c | 313.08b | 261.72c | 275.51c | 68.71 |
| Shank (g) | 107.42b | 104.03b | 87.97c | 221.3a | 95.97c | 128.34b | 114.30b | 105.31b | 6.13 |
| Wing (g) | 184.95b | 208.21a | 117.67b | 233.83b | 178.10b | 213.77a | 189.71b | 188.40b | 4.69 |
| Head (g) | 70.27 | 73.24 | 62.98 | 79.83 | 64.80 | 76.53 | 69.53 | 70.88NS | 2.03 |
| Neck (g) | 123.03 | 117.41 | 106.38 | 137.83 | 118.20 | 122.81 | 113.09 | 133.91NS | 2.46 |

 |  |  |

 Means within a row with different superscripts differ (P<0.05) significantly.

The result of the effect of substitution of antibiotics with selected medicinal plant leaf meals on the carcass characteristics indicated that the dressed weight was highest for T4 (2052) g followed by T6 (1772) g, T8 (1763) g, T7 (1709) g, T2 (1666.67) g, T5 (1631) g, T1 (1627) g and T3 (1557.33) g respectively. This result showed that medicinal plant leaf meals such as *Spondias mombin* or a combination of medicinal plant leaf meals can replace antibiotics in poultry production. The dressed weight of T4 was significantly (P<0.05) higher than the other treatment groups.

 The result also showed that the birds fed on diets containing *Spondias mombin* leaf meals performed better than other leaf meal combinations. Asuquo *et al*. (2013) reported a significant reduction (P<0.05) in the hormonal levels such as follicle-stimulating hormone (FSH), luteinizing hormone (LH), oestradiol and progesterone in female Wistar rats when administered with *Spondias mombin* leaf extract. These reductions must have led to the broilers developing masculinity, thus enhancing weight gain.

The dressing percentage was also significantly (P<0.05) higher for T4 than other treatment groups. The breast muscle cut, thigh, drumstick, shank, and wing were significantly (P<0.05) higher for T4 when compared to other treatment groups. The head and neck, however, did not differ (P>0.05) significantly. This agrees with the result of Windisch (2009) and Guo *et al*., (2004) that phytogenic feed additive improves carcass quality.

Effects of substitution of antibiotics with selected medicinal plant leaf meals on the various

 organs of broiler chicken are presented below in Table 6:

**Table 6: Effect of the selected medicinal plant leaf meals on organ characteristics of broiler chicken**

|  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  **TREATMENT**

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Parameters** |  **1** |  **2** |  **3** |  **4** |  **5** |  **6** |  **7** |  **8** | **SEM +** |
| Empty gizzard (g) | 49.30 | 52.53 | 51.32 | 53.35 | 45.03 | 46.00 | 45.53 | 51.16NS | 4.63 |
| Crop (g) | 22.02 | 20.70 | 22.50 | 21.48 | 18.39 | 2183 | 21.31 | 19.56NS | 168 |
| Proventicles (g) | 19.77 | 20.06 | 15.19 | 23.45 | 18.46 | 21.45 | 19.27 | 20.72NS | 1.92 |
| Small intestine (g) | 66.38 | 64.82 | 61.10 | 73.74 | 68.82 | 72.58 | 67.31 | 64.63NS | 0.83 |
| Large Intestine (g) | 42.87 | 42.73 | 38.53 | 47.59 | 40.46 | 44.38 | 41.67 | 41.88NS | 2.83 |
| Caecum (g) | 14.62 | 15.43 | 13.24 | 15.40 | 13.20 | 16.37 | 14.35 | 14.36NS | 1.37 |
| Heart (g) | 13.26b | 12.36b | 14.32b | 21.75a | 22.00a | 21.16a | 12.18b | 14.57b | 4.31 |
| Abdominal fat | 23.69b | 23.68b | 13.00a | 22.38b | 21.97b | 23.51b | 18.25a | 18.55a | 3.72 |
|  |  |  |  |  |  |  |  |  |  |

 |  |  |

 Means within a row with different superscripts differ (P<0.05) significantly.

The organs of the broiler chicken such as gizzard, crops, proventriculus, small intestine, large intestine and caecum showed no significant (P>0.05) differences between the treatment groups.

However, the heart of the broiler birds from T4 was significantly (P<0.05) heavy than those of the treatment groups. The abdominal fats were significantly (P<0.05) lower in T3, T7 and T8 than in other treatment groups. This may be due to the anti-adipogenic effect of various leaf meals, which inhibits the accumulation of lipid droplets in the fat cell (Asuquo *et al*., 2013)

Effects of substitution of antibiotics with selected medicinal plant leaf meals on

 haematological parameter of broiler chicken is presented in Table 7:

**Table 7:** **Effects of substitution of antibiotics with selected medicinal plant leaf**

 **meals on the haematological parameter of broiler chicken**

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Parameters** |  **T1** | **T 2** |  **T3** | **T4** | **T5** | **T6** | **T7** | **T8** | **SEM +** |
| WBC (x103/ul) | 6.63c | 7.34c | 17.73a | 11.43b | 6.98c | 11.34b | 7.36c | 7.54c | 4.21 |
| Hb (g/dl) | 109.00b | 119.34b | 102.67b | 112.00b | 110.00b | 112.67b | 130a | 121ab | 3.05 |
| RBC (x106/ul) | 2.38 | 2.57 | 2.29 | 2.56 | 2.44 | 2.48 | 2.79 | 2.67NS | 0.62 |
| MCV (fl) | 136.90 | 139.50 | 136.67 | 130.40 | 133.60 | 128.67 | 128.65 | 135.5NS | 1.27 |
| Platelets (x103/ul) | 21.33a | 13.67b | 6.33c | 10.00b | 10.0b | 7.33c | 6.67c | 7.67c | 1.21 |
| MCH (pg) | 45.50 | 44.40 | 44.76 | 45.80 | 44.37 | 45.40 | 42.93 | 45.10NS | 1.75 |
| MCHC (g/dl) | 326.67 | 332.67 | 334 | 335.67 | 337.33 | 328.64 | 332.34 | 333.34NS | 2.26 |

Means within a row with different superscripts differ (P<0.05) significantly.

The result of the effect of the substitution of antibiotics with selected medical plant leaf meals on the haematological particles indicated that the white blood cell count was highest for T3 (17.73) followed by T4 (11.43), T6 (11.34) T8 (7.54), T7 (7.36), T2 (7.34), T5 (6.98) T1 (6.63) respectively. This result shows that broilers fed neem leaf meals (T3) had significantly (P<0.05) increased white blood cells when compared to broilers in the other treatment groups, while T1, T2, T5, T7 and T8 did not differ significantly (P>0.05) from each other. The high white blood cell count in broilers that received neem is due to its secondary metabolic like az*adirachtin*, *Nimbin*, *salain*, *meliacin* and their natural derivatives (NRC,1995). Neem has also been associated with an immune response by activating macrophages and lymphocytes in animals. Neem also processes a range of other pharmacological properties, such as being anti-inflammatory, anti-hyperglycaemic, anti-ulcers, immunomodulatory and various other properties showing no adverse effects (Gannu *et al.*, 2003; Chakraberty, 2012).

The haemoglobin count of the broilers finisher in T7 was significantly (P<0.03) higher than in the other treatment groups. Their red blood cell count and the mean cell volume (MCV) showed no significant (P>0.05) differences among the treatment group. The effects of the medicinal plant leaf meals on platelets indicated that T1 was significantly (P<0.05) higher when compared to the other treatment groups, while T3 (6.33x103), T7 (6.67 x103), T6 (7.33 x103) and T8 (7.67 x103) were significantly (P>0.05) lower. The MCH and MCHC values for the groups showed no significant (P>0.05) differences among the treatment groups. This can be attributed to the antioxidant capacities of the test ingredient. Red blood cell is formed in the long bones of the body, and sufficient production is dependent on the amount of iron absorbed from food digested. Furthermore, the key constituent of erythrocytes is haemoglobin, as it forms about one-third of red blood cell content.

However, the values achieved in all haematological analyses fall into the acceptable range for clinically healthy chicken (Mitruka and Rawnlay, 1981).

Conclusively, given the economic importance of poultry as a source of food, money, and employment most farmer is desirous of improved fed conversion ratio, dressed weight and dressed percentage. 5 percent inclusion of *Spondia mombin;* 2.5 percent combination each of neem and *Spondia* and 1.66 percent combination each of *Moringa oleifera*, neem, and *Spondia mombin* respectively is thus recommended

 **References**

Adedeji, O.S Ferinu, G.O. S. A. Amen, and T. B. Olayemi (2008). The effect of dietary

Bitter Kola (Garcinia Kola inclusion on Body weight haematology and Survival Rate of

Pullet chicks, Journal of Animal and Veterinary Advances; 5(3), 184-187.

Alaku, S.O. 2010. Introduction to Animal Science Jee Communications. Enugu. Pp 88.

Consumer Updates. 2013. Phasing out certain Antibiotic use in farm Animals.

[Http://www.fad.gov/forconsumers/consumersupdates/ucm378100.thm](http://www.fad.gov/forconsumers/consumersupdates/ucm378100.thm)

Dilger, R.N and baker, D.H. 2008. Cysteine imbalance and its effect on

precursor acclimatization in chicks. Journal of Animal Science, 86: 1832-1840.

Iji, P.A. 2009. The impact of cereal anti-nutritive factors on intestinal development and

function in broiler chickens. *World Journal of poultry science;* 55:375-387.

Iji, P.A., M.M. Bhuiyan, M.R. Barakatain, N. Chaugnarong, N and A. P.Widodo. 2011.

Improving the Nutritive Value of Alternative Feed Ingredients for Poultry. University of

New England, Engormix. Com.

Gefu, J.O and C.B. Alawa. 2000. Ethno vet practices research Development process of

the workshop on ethno vey practices held 14-18 August 200, Kaduna, Nigeria.

Published by the NAPRI, ABU, Zaria. Nigeria.

Olukosi, O.A., Gowieson, A.J., Adeola O. (2010). Broiler responses to supplementation

of phytase and admixture of carbohydrates and proteases in maize, soya bean meal

diets with or without maize DDGS with solubles. *British* *Journal of Poultry Science;* 51:

434-443.

Maigandi, S. A and M. K. Usman. 1996. A survey of turkey production in Sokoto state

Nigeria. ANPRD Newsletter. 6:5-7.

Nwude, N., and Ibrahim. M.A. 1980. Plants used in traditional veterinary Medicinal

practice.

Oyekunle, M.A and M. O. Owonikoko. 2002. Antimicrobial drug usage for poultry

production within a local government area in Ogun State, *Nigeria Journal of Animal*

*Production;* 29(1): 113-120.

Ugwu, L.L C 2006. Animal feeds and feeding staff. Lecture notes, unpublished. Pp. 26-

30.

Sonaiya, E.B. 1993. Evolution of Non-Conventional feed ingredients as supplements

for scavenging chicken. Proceeding vii world conference on Animal Production.

Edmonton. Canada. Pp. 28-29.

FAO., 2008. Food and Agricultura Organization of United National: Economic and Social

Department: The Statistical Division. Top ten Ginger producers.

Abaza, I. M. K.(2001. Using some medicinal plants as feed additives in broiler diets. *Life*

*Science Journal* 11: 240 – 248.

Anikwe, M. A. N., J.C. Eze. M.C. Chima and E.E. Ikenganyia. 2016. Soil physicochemical

quality in contrasting tillage systems and its effect on nodulation and nodulation

effectivity of groundnut, Bambara groundnut and soybean in a degraded Ultisol in

Agbani, Enugu Southeastern Nigeria. *The rhizosphere*, *1*, 14-16.

Anikwe, M. A. N., E. E. Ikenganyia. J. Egbonimale and C. Oputah. 2017. Assessment of

some tropical plants for the phytoremediation of petroleum-contaminated soil: Effects of

remediation on soil physical and chemical properties. *International Journal of Plant and*

*Soil Science*, *14*(2), 1-9.

Asuquo, O.R.E., S. Brownson. G. B. Umuetuk and I. S. Utin. 2013. Effects of ethanolic

leaf extract on Spondias mombin on the pituitary-gonadal axis of female Wistar rats.

*International Journal of Science and Research,* vol. 2, No. 9, pp.5-8.

Chakraberty, R. A. 2012. Sacred trees of temples of Tiruchirappalli, Tamil Nadu-The

Natural and Ecological Heritage of India. *Global Journal of Research on Medicinal*

*Plants & Indigenous Medicine*, *1*(6), 225.

Bengmark, S.1998. Ecological control of the gastrointestinal tract. The role of probiotic

flora. Gut, 42: 2-7.

Dhama, K., V. Verma. V. P.M. Sawant. R. Tiwari. R. K. Vaid and R. S. Chauhan. 2011.

Applications of probiotics in poultry: Enhancing immunity and beneficial effects on

production performances and health: A review. *J. Immunol. Immunopathol*, *13*(1), 1-19.

Frankic, T., M. Voljg, J. Salobir. J and V. Rezar. 2009. Importance of Medicinal Herbs in

Animal Feeding; A Review; *Acta Agric, Slovenia,*92:95-96.

Gill, D. M. 2001. ADP-ribosylation of p21ras and related proteins by Pseudomonas

aerurinosa exoenzyme S. *Infection and immunity*, *59*(11), 4259-4262.

Gannu, R., S. K.Yamsani, A. Devandla, V. V. Yamsani, B. Athukuri, C.R. Palem and

S.Manda. 2003. Effect of ashwagandha and aloe vera pretreatment on intestinal

transport of buspirone across rat intestine. *Latin American Journal of Pharmacy*, *30*.

Guo, F.C., R.P. Kwakkel, D. S. Williams and M. N. Li. 2004. Effects of mushroom and

herb polysaccharides, as alternatives for an antibiotic, on growth performance of broilers.

*Br. Poultry Sci*., 45: 684-694.

Hassan, H. M.A., A. W. Mohamed, E. R . Youssef and B. F. Hassan. 2010. Effect of

using organic acids to substitute antibiotic growth promoters on performance and

intestinal microform of broilers. *Asian-Aust. J. Anim. Sci*., 23: 1348-1353.

Isman, M. B., Y. Akhtar, V. V. Pathak and V. Kumar. 1991. Comparative growth

inhibitory and antifeedant effects of plant extracts and pure allelochemicals on four

phytophagous insect species. *Journal of Applied Entomology*, *128*(1), 32-38.

Mitruka, B.M. and A. M Rawnsley. 1981. Clinical biochemical and haematological

references values in normal experimental animals and normal humans. New York:

Masson Pub.USA

Nava, G.M., H. R. Attene-Ramos, R. D. Gaskins and J. D. Richards. 2009. Molecular

analysis of microbial community structure in the chicken ileum following organic acid

supplementation. *Vet Microbiol*., 137: 345-353.

Overland, M., N. P. Granli, O. Kjos, S. H. Fjetland, M. Steien and M. F. Stokstad 2000.

Effect of dietary formates on growth performance, carcass traits, sensory quality,

intestinal microflora and stomach alterations in growing-finishing pigs. *Int. J. Anim*. Sci.,

78: 1875-1884.

Seema, A. and T. S Johri. 1992. Probiotics in poultry feed. *Poult. Guide*, *29*(1), 51-58.

SPSS, 2012. IBM SPSS statistics version 22. *Boston, Mass: International Business*

*Machines Corp*, *126*.

Snyder, I.L, and B. S. Wostmann. 1987. The growth rate of male germ-free Wistar rats

fed ad libitum or restricted natural ingredient diet Lab. *Anim. Sci*., 37: 320-325.

Windisch, W., K. Schedle, A. Plitzner and A. B. Krcismayr. 2009. Use of phytogenic

products as feed additives for swine and poultry. *J. Anim. Sci*., 86; 40- 48.