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# MIEN OF RETINOBLASTOMA PROTEIN (pRb) IN GRADES OF CERVICAL LESION

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### ABSTRACT

The retinoblastoma protein (pRb); gene appellation (shortened RB or RB1) is a tumor suppressor protein that stands dysfunctional in numerous major malignancies. Cervical cancer is another utmost common cause of cancer-related death globally between women folk. This study was aimed at investigating the expression of retinoblastoma protein (pRb) in grades of cervical lesion. Two hundred (200) cases of archived formalin fixed paraffin embedded (FFPE) tissues of already diagnosed cervical lesions and cervical carcinoma tissue blocks were selected for this study. Sections of 3µ thickness were cut for immunohistochemical staining technique. The immunohistochemical staining was semi-quantitatively scored based on percentage of cells that stained positive and the intensity of the staining. Photomicrograph was basically used for analyzing the expression, and comparative analysis of the data generated from the expression was statistically done. Frequency of data was calculated using Chi-square  $(x^2)$ . The expression of retinoblastoma protein (pRb) was found to be 95.3% in cervical intraepithelial neoplasia grade I (CIN I) cases, 100% in CIN II cases, 85.4% in CIN III cases, and 60.8% in squamous cell carcinoma (SCC) cases. The expression of pRb in CIN I, II, and III as well as SCC significantly reduced as the CIN cases matured to malignancy (p<0.05) with CIN III been observed with the least expression of pRb among CIN cases. The downregulation of pRb observed in this study may indicate mutation of RB gene in cervical lesions and could be involved in cervical carcinogenesis.

Keywords: Cervical, dysplasia, immunoreactivity, retinoblastoma, protein.

## **INTRODUCTION**

The retinoblastoma (RB) tumor suppressor protein plays a pivotal role in the control of cell cycle, terminal differentiation, and various other biological events (Dyson, 2016). RB is genetically or functionally inactivated in numerous human cancers, including retinoblastoma, small cell lung cancer (SCLC), prostate cancer, and breast cancer (Shunsuke et al. 2020). The retinoblastoma susceptibility gene (RB1) was the first tumor suppressor gene to be molecularly defined and the retinoblastoma protein (pRb) which is its gene product regulates transcription and is a negative regulator of cell proliferation (Dyson, 2016). The canonical pathway whereby RB exerts its tumor suppressive activity entails the formation of a transcriptional repression complex with E2F transcription factors and

various chromatin modifiers, such as histone deacetylases (HDACs). This complex orchestrates the G1/S transition during cell cycle progression primarily by controlling E2F target genes (Dyson, 2016; Kitajima and Takahashi, 2017).

The Rb1-encoded protein (pRb) is well known as a general cell cycle regulator, and this activity is critical for pRb-mediated tumor suppression (Goodrich, 2006). Better insight into pRb-mediated tumor suppression and clinical exploitation of pRb as a therapeutic target will require a global view of the complex interdependent network of pocket protein complexes that function simultaneously within given tissues (Goodrich, 2006). Most mitogenic signals commonly merge on the transcriptional up regulation of D-type cyclins and then stimulate cyclin-dependent kinases, including CDK4/6. D-type cyclin-CDK4/6 complexes

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have been proposed to promote monophosphorylation on RB, which allow it to exert early G1 functions by starting the release of E2Fs. E2F target genes, including cyclin E and A, in cooperation with CDK2 or CDK1, are responsible for full phosphorylation of RB at 13 remaining sites. This allows cells to enter the S and M phases (Rubin, 2013; Dyson, 2016; Sanidas et al. 2019). The fact that genetic and/or epigenetic aberrations of the components in the RB pathway tend to be mutually exclusive in the patients might implicate a linearity of the RB pathway (Shunsuke et al. 2020).

Cervical cancer is a major gynaecological cancer which encompasses hysterical cell division and tissue invasiveness of the female uterine cervix (Dasari et al. 2015). Cervical cancer is the second most common cause of cancer-related death among women worldwide, with over 500,000 new cases diagnosed annually and 50% mortality rate in Asia (Daniyal et al. 2015). Cancer cells can disrupt the immune response through the over expression of inhibitory molecules like PD-L1 (Patel and Kurzrock, 2015) or the loss of expression of stimulatory molecules like CD40L. Tumors can evade immune surveillance even in the presence of tumor antigens because the immune cells may not receive adequate signals for activation and proliferation or are suppressed by inhibitory checkpoint proteins (Whiteside, 2006). The decision to commit to a new round of cell division occurs when the cell activates cyclin-CDK-dependent transcription which promotes entry into S phase (Bertoli et al.2013). During early G1, the transcriptional repressors Rb (retinoblastoma), p107 and p130, known as pocket proteins, bind to the E2F transcription factors to prevent G1-to-S transition. Rb binds and represses activator E2F transcription factors (E2F1-3), while p107 and p130 bind E2F4 and E2F5 respectively to form complexes which repress transcription of G1-to-S promoting factors (proteins) (Bertoli et al. 2013). This study therefore investigated the expression of retinoblastoma protein (**pRb**) in grades of cervical lesion.

# **MATERIALS AND METHODS Study Population**

This study was conducted on tissue blocks from women that had CIN I, II, III and cancer of

the cervix in Lagos University Teaching Hospital, Lagos. The age of the patients ranged from 28 to 64 years. The duration of the tissue blocks used was from 2002-2016.

### **Ethical Consideration**

Ethical approval for this study was obtained from the Ethics and Research Committees of Lagos University Teaching Hospital, Lagos

### **Specimen collection**

Two hundred (200) cases of archived formalin fixed paraffin embedded (FFPE) tissues of already diagnosed cervical lesions and cervical carcinoma tissue blocks were selected for this study.

#### **Histopathological Procedures**

Each formalin fixed paraffin wax embedded tissue was sectioned to harvest tissue areas with the highest lesion. Sections of 3u thick were cut for immunohistochemical staining technique. Four sections were obtained from each block of cervical intraepithelial neoplasia grade I (CIN I), CIN II, CIN III and cervical carcinoma; from which one section was used for haematoxylin and eosin staining technique, one section was treated with retinoblastoma protein (pRb) while the other section served as negative and positive controls. Haematoxylin and eosin method was carried out to confirm earlier diagnosis before proceeding to the immunohistochemical analysis.

Haematoxylin and Eosin Staining Technique (Avwioro, 2014) was used to stain the sections and Immunohistochemical Technique was performed (Marc, 2009).

# Control for Immunohistochemistry (IHC):

Positive control sections for the marker was obtained from tissues that are known to express the antigen. In negative control, tissues that are kown not to express the antibody marker was used, while in the reagent negative controls, pRb antibody that is being tested was omitted.

## **Reading of Immunohistochemistry results**

Cells with specific brown colours in the cytoplasm, cell membrane or nuclei depending on the antigenic sites were observed. The haematoxylin stained cells without any form of An Official Publication of Enugu State University of Science & Technology ISSN: (Print) 2315-9650 ISSN: (Online) 2502-0524 2

### RESULTS

brown colours were scored negative. Nonspecific binding/brown artifacts on cells and connective tissue was disregarded (Marc, 2009). Immunoreactive Scoring System by Klein et al. (1999) was used.

# **Statistical Analysis**

Photomicrograph was basically used for analyzing the expression and comparative analysis of the data generated from the expression of the pRb protein were statistically analyzed by the statistical data software SPSS version 16 (SPSS Inc. Chicago, Illinois) for windows. Descriptive statistical analysis was also used to analyze some of the data.

The tables below summarize the expression of retinoblastoma protein (pRb) in cervical intraepithelial neoplasia grade I (CIN I), CIN II, CIN III, and Cervical squamous cell carcinoma (SCC).

Retinoblastoma protein (pRb) was highly expressed in 48.9% of the overall CIN diagnosis but in 10.1% of SCC cases. Low expression of pRb was observed in 46.5% and 50.7% of overall CIN and SCC cases respectively. pRb expression was significantly reduced as the dysplasia progresses (p<0.05) (Table 1).

Case	No.	Positive E	xpression	Negative Expression
	Tested	High(%)	Low(%)	(No expression)(%)
CIN	131	64(48.9)	59(46.5)	8(4.6)
SCC	69	7(10.1)	35(50.7)	27(39.2)
				*p<0.05

Table 1: Expression of pRb in Overall CIN and SCC

pRb was most expressed among participants diagnosed of CIN II (53.2%), followed by CIN I(48.8%), CIN III (43.9%) whereas those diagnosed of SCC had the least expression of pRb (10.1%). The expression of pRb in CIN I, II, and III as well as SCC

significantly reduced as the CIN cases matured to malignancy (p<0.05) with CIN III been observed with the least expression of pRb among CIN cases. Generally, the SCC revealed the least expression of pRb (10.1%) in all dysplasia cases (Table 2).

Table 2: Expression of pRb in CIN I, CIN II, CIN III, and Squamous cell carcinoma of the Cervix

Case	No.	<b>Positive Expression</b>		Negative Expression
	Tested	High (%)	Low (%)	(No expression)(%)
CIN I	43	20(46.5)	21(48.8)	2(4.7)
CIN II	47	22(46.8)	25(53.2)	0
CIN III	41	17(41.5)	18(43.9)	6(14.6)
SCC	69	35(50.7)	7(10.1)	27(39.2)
CIN - Cervical Intraepith	*p<0.05			

SCC - Squamous cell carcinoma of the Cervix.

In this study, strong expression of pRb was observed in CIN I which was demonstrated endogenously with the brownish pigmentation on the section (Fig. 1).

Moderate expression of pRb was observed in CIN II that was demonstrated endogenously with the brownish pigmentation on the section (Fig. 2).

Mild expression of pRb was observed in CIN III which was demonstrated endogenously with the brownish pigmentation on the section (Fig. 3).

Low expression of pRb was indicated in SCC which was demonstrated endogenously with the brownish pigmentation on the section (Fig. 4).

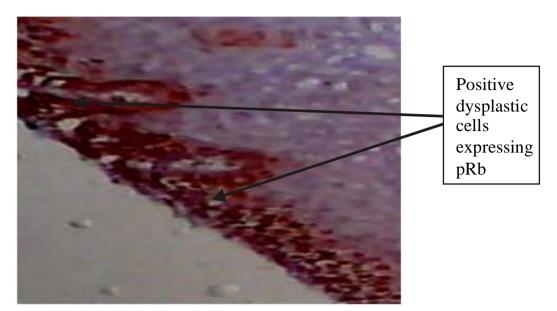


Fig. 1: Immunohistochemistry of Strong Expression of pRb in CIN I (mag x400)

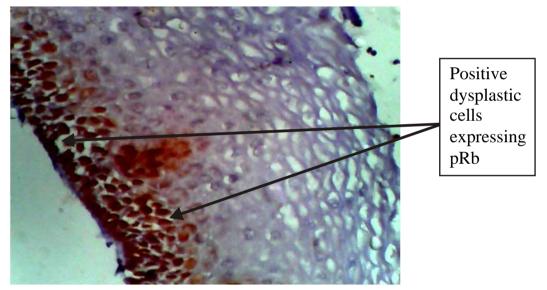


Fig. 2: Immunohistochemistry of Moderate Expression of pRb antibody in CIN II (mag x400)

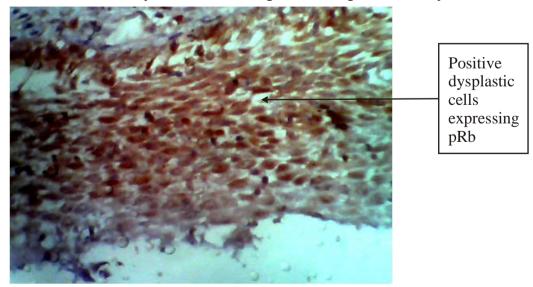
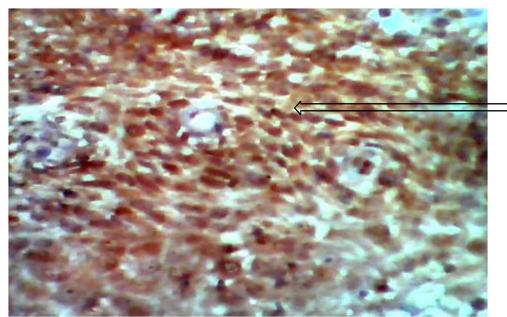


Fig. 3: Immunohistochemistry of Mild Expression of pRb antibody in CIN III (mag x400)



Dysplastic cells expressing pRb

Fig. 4: Immunohistochemistry of Low Expression of pRb antibody in SCC (mag x400)

# DISCUSSION

The RB tumor suppressor pathway has been extensively studied (Dick et al. 2018). The retinoblastoma protein (pRb) is a cell cycle regulator predominantly known as cell cycle repressor deactivated in most human cancers. Loss of pRb function results from mutations in the gene coding for pRb or for any of its upstream regulators (Sosa-García et al. 2010). The inactivation of RB1 through somatic mutation, deletion, or epigenetic silencing in various cancers has been reported. Hypophosphorylated pRB binds to and represses the transcriptional activity of E2F family members, which control the expression of genes necessary for cell cycle progression (Wang et al. 2018).

In this study, the expression of retinoblastoma Protein (pRb) was observed to be 95.3% in CIN I cases, 100% in CIN II cases, 85.4% in CIN III cases, and 60.8% in SCC cases. Low level expression of pRb was observed to be more frequent in SCC cases (50.7%) followed by CIN II (46.8%), CIN I (46.5%), and CIN III (41.5%). This study showed pRb immunoreactivity to be frequent in majority of the CIN and SCC cases which is comparable to the findings by several authors that reported ubiquitous expression of pRb in normal or cancerous tissues (Horowitz et al. 1990; Furukawa et al. 1991; Cordon-Cardo and

Richon, 1994; Salcedo et al. 2002); Some studies that were also done on normal uterinecervix tissue shows that pRb is expressed in mature and differentiated cells, in the basal third epithelium in 90% of normal/reactive atypia or in the scattered nuclei of normal cells in all cases tested (Benedict et al. 1990; Cance et al. 1990; Cordon-Cardo et al. 1992).

This study observed that pRb immunoreactivity in SCC was frequently lower than in CIN which is in agreement with the report of some workers (Ludlow et al. 1993; Boyer et al. 1996) suggesting that the down regulation of retinoblastoma (RB) gene could be involved in cervical carcinogenesis. The low expression of pRb may also be linked to inactivation of pRb which is as a result of complex formation with high risk HPV E7 oncoprotein and its degradation, down regulation mechanisms (Boyer et al. 1996; Whyte et al. 1988; Dyson et al. 1989). The heterogeneous pRb immunostaining observed in CIN stages as compared to SCC in this study is similar to that reported by Salcedo et al. (2002) which also observed heterogeneous pRb immunostaining during the different stages of cervical carcinogenesis and suggest that this staining pattern could be a common feature implicated in pathological process of uterinecervix carcinoma. This study also showed a positive statistical correlation between the degree of dysplasia and the degree of pRb expression in all the cases which indicates that as degree of cervical dysplasia increases (from premalignant to malignant lesion), there will be gradual decrease in pRb expression; this findings on CIN and SCC can be related to the suggestion of some authors that pRb negative tumours might be clinically more aggressive and with a poorer prognosis than those tumours containing a variable pRb expression (Benedict et al. 1990; Hanson et al. 1994; Harbour et al. 1988). In support of this possibility, some studies also reported that high RB gene expression inhibits tumour cell invasion *in vitro* (Li et al. 1996).

## **CONCLUSION**

Retinoblastoma protein (pRb) expression revealed a progress in the degree of cervical dysplasia (premalignant to malignant lesion), with a corresponding gradual decrease in pRb expression. The pRb immunoreactivity in SCC is frequently lower than the overall CIN cases. The down regulation of retinoblastoma (RB) protein may indicate mutation of RB gene in cervical lesions and it could be involved in cervical carcinogenesis.

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## **CONFLICT OF INTEREST**

Authors declare no conflicts of interest

## REFERENCES

- <u>Ana MA, Howard MS.</u> (2015). Tumor-suppressor genes, cell cycle regulatory checkpoints, and the skin. North American Journal of Medical Sciences. 7(5): 176–188.
- Avwioro OG. (2014). Histochemistry and Tissue Pathology in; Principle and Technique. Third edition, Claverianun Press, Ibadan, Nigeria. pp 133-168.

- Benedict WF, Xu HJ, Hu SX, Takahashi R. (1990). Role of the retinoblastoma gene in the initiation and progression of human cancer.Journal of Clinical Investigations. 85(4):988-993.
- Bertoli C, Skotheim JM, de Bruin, RAM. (2013). Control of cell cycle transcription during G1 and S phases. Nature Reviews Molecular Cell Biology. 14(8): 518–528.
- Boyer SN, Wazer DE, Band V. (1996). E7 protein of human papilloma virus-16 induces degradation of retinoblastoma protein through the ubiquitinproteasome pathway. Cancer Research. 56(20):4620-4624.
- Cance WG, Brennan, MF, Dudas ME, Huang CM, Cordon-Cardo C. (1990). Altered expression of the retinoblastoma gene product in human sarcomas. The New England Journal of Medicine. 323(21):1457-1462.
- Collins M, Ling V, Carreno BM. (2005). The B7 family of immune-regulatory ligands. Genome Biology.6:223
- Cordon-Cardo C, Richon VM. (1994). Expression of the retinoblastoma protein is regulated in normal human tissues. American Journal of Pathology. 144(3):500-510.
- Cordon-Cardo C, Wartinger D, Petrylak D, Dalbagni G, Fair WR, Fuks Z, Reuter VE. (1992). Altered expression of the retinoblastoma gene product: prognostic indicator in bladder cancer. Journal of the National Cancer Institute. 84(16):1251-1256.
- Daniyal M, Akhtar N, Ahmad S, Fatima U, Akram M, Asif HM. (2015). Update knowledge on cervical cancer incidence and prevalence in Asia. Asian Pacific Journal of Cancer Prevention. 16;(9): 3617-3620.
- Dasari S, Wudayagiri R, Valluru L. (2015). Cervical cancer: Biomarkers for diagnosis and treatment. Clinica Chimica Acta. 445: 7-11.
- Dick FA, Goodrich DW, Sage J, Dyson NJ. (2018). Noncanonical functions of the RB protein in cancer. Nature Reviews Cancer 18: 442–451.
- Dyson N, Howley PM, Münger K, Harlow E. (1989). The human papilloma virus-16 E7 oncoprotein is able to bind to the retinoblastoma gene product. Science. 243(4893):934-937.
- Dyson NJ. (2016). RB1-A prototype tumor suppressor and an enigma. Genes and Development. 30:1492–1502.

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- Engel BE, Cress WD, Santiago-Cardona PG. (2015).The Retinoblastoma Protein: A master tumor suppressor acts as a Link between cell cycle and cell adhesion. Cell Health Cytoskeleton. 7: 1–10
- Furukawa Y, DeCaprio JA, Belvin M, Griffin JD. (1991). Heterogeneous expression of the product of the retinoblastoma susceptibility gene in primary human leukemia cells. Oncogene. 6(8):1343-1346.
- Goodrich DW. (2006): The retinoblastoma tumorsuppressor gene, the exception that proves the rule. Oncogene.25:5233-5243.
- Hanson KD, Shichiri M, Follansbee MR, Sedivy JM. (1994). Effects of c-myc expression on cell cycle progression. Molecular and Cell Biology. 14(9):5748-5755.
- Harbour JW, Lai SL, Whang-Peng J, Gazdar, AF, Minna JD, Kaye FJ. (1988). Abnormalities in structure and expression of the human retinoblastoma gene in SCLC. Science. 241(4863):353-357.
- Horowitz JM, Park SH, Bogenmann E, Cheng JC, Yandell DW, Kaye FJ, Minna JD, Dryja TP, Weinberg RA. (1990). Frequent inactivation of the retinoblastoma anti-oncogene is restricted to a subset of human tumor cells. Proceedings of the National Academy of Sciences of the United States of America. 87(7):2775-2779.
- Kitajima S, Takahashi C. (2017). Intersection of retinoblastoma tumor suppressor function, stem cells, metabolism, and inflammation. Cancer Science. 108:1726-1731.
- Klein M, Picard E, Vignaud JM, Marie B, Bresler L, Toussaint B, Weryha G, Duprez A, Leclère J. (1999). Vascular endothelial growth factor gene and protein: strong expression in thyroiditis and thyroid carcinoma. Journal of Endocrinology. 161(1):41-49.
- Korenjak M, Brehm A. (2005). E2F-Rb complexes regulating transcription of genes important for differentiation and development. Current Opinion in Genetics & Development. 15 (5): 520-527.
- Li J, Hu SX, Perng GS, Zhou Y, Xu K, Zhang C, Seigne J, Benedict WF, Xu HJ. (1996). Expression of the retinoblastoma (RB) tumor suppressor gene inhibits tumor cell invasion in vitro.Oncogene. 13(11):2379-2386.
- Locksley RM, Killeen N, Lenardo MJ. (2001). The TNF and TNF receptor superfamilies: integrating mammalian biology. Cell. 104(4):487-501.

- Ludlow JW, Glendening CL, Livingston DM, DeCarprio JA. (1993). Specific enzymatic dephosphorylation of the retinoblastoma protein. Molecular and Cell Biology. 13(1):367-372.
- Marc M. (2009). Standard Immunohistochemistry Staining Method; Avidin Biotin Complex (ABC) Method. IHC world life science products and services. http://www.ihcworld.com/\_protocols/general\_I HC/standard\_abc\_method.htm (Accessed on 20th June, 2018).
- Mohammed A, Ahmed SA, Oluwole OP, Avidime S. (2006). Malignant tumours of the female genital tract in Zaria, Nigeria: Analysis of 513 cases. Annals of African Medicine. 5:93-96.
- Rubin SM. (2013). Deciphering the retinoblastoma protein phosphorylation code. Trends in Biochemical. Sciences. 38:12-19.
- Salcedo M, Taja L, Utrera D, Chávez P, Hidalgo A, Pérez C, Benítez L, Castañeda C, Delgado R, Gariglio P. (2002). Changes in retinoblastoma gene expression during cervical cancer progression. International Journal of Experimental Pathology. 83(6):275-286.
- Sanidas I, Morris R, Fella KA, Rumde PH, Boukhali M, Tai EC, Ting DT, Lawrence MS, Haas W, Dyson NJ. (2019). A Code of Mono-phosphorylation modulates the Function of RB. Molecular Cell Journal. 73:985-1000.
- Shunsuke K, Fengkai L, Chiaki T. (2020). Tumor Milieu Controlled by RB Tumor Suppressor. International Journal of Molecular Sciences. 21:2450
- Wang LH, Wu CF, Rajasekaran NB, Shin YK. (2018). Loss of Tumor Suppressor Gene Function in Human Cancer: An Overview. Cellular Physiology and Biochemistry. 51:2647-2693
- Whiteside TL. (2006). Immune suppression in cancer: effects on immune cells, mechanisms and future therapeutic intervention. Semin Cancer Biology.16:3-15.
- Whyte P, Buchkovich KJ, Horowitz JM, Friend SH, Raybuck M, Weinberg RA, Harlow E. (1988). Association between an oncogene and an antioncogene: the adenovirus E1A proteins bind to the retinoblastoma gene product. Nature. 334(6178):124-129.

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